

1 **Environmental gradients decouple demographic and adaptive connectivity**
2 **in a highly mobile coastal marine species**

3 Chris Brauer¹, Andrea Bertram¹, Jonathan Sandoval-Castillo¹, Anthony Fowler², Justin Bell³,
4 Paul Hamer^{3,4}, Maren Wellenreuther^{5,6}, Luciano B. Beheregaray¹

5

6 ¹Molecular Ecology Laboratory, College of Science and Engineering, Flinders University, Bedford
7 Park, SA, Australia

8 ²Aquatic Sciences, South Australian Research and Development Institute, Henley Beach, SA,
9 Australia

10 ³Victorian Fisheries Authority, Queenscliff, Vic, Australia

11 ⁴Pacific Community, Noumea, New Caledonia

12

13 ⁵The New Zealand Institute for Plant and Food Research Limited, Nelson, New Zealand

14 ⁶The School of Biological Sciences, University of Auckland, Auckland, New Zealand

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17 Corresponding author: tel: +61 8 8201 5243; email: luciano.beheregaray@flinders.edu.au

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23 **Abstract**

24 Understanding how eco-evolutionary processes shape genetic variation and persistence in
25 marine species with highly variable recruitment dynamics and dispersal potential remains a
26 fundamental challenge, particularly when considering the interplay between gene flow and
27 local adaptation. Here, we employed a seascape genomics approach to investigate population
28 connectivity and local adaptation in Australasian snapper (*Chrysophrys auratus*, Sparidae)
29 along 1500 km of the environmentally heterogeneous southern Australian coastline. Using
30 14,699 SNPs, we identified two distinct regional populations aligned with known
31 biogeographical regions. Genotype-environment association analyses revealed 855 candidate
32 adaptive loci associated with environmental variation, including temperature, salinity, and
33 primary productivity. Connectivity analyses using neutral markers indicated high gene flow
34 throughout both eastern and western regions, while candidate adaptive loci revealed
35 substantially reduced connectivity, especially for Northern Spencer Gulf and West Coast
36 populations. This pattern of locally restricted gene flow at adaptive genomic regions suggests
37 that strong environmental gradients are driving adaptive divergence despite high overall
38 connectivity. Our results support contingent migration as a potential mechanism modulating
39 the balance between local adaptation and gene flow in this economically and ecologically
40 important marine species. These findings might also have implications for regional
41 management of other coastal fisheries that are experiencing substantial declines. As climate
42 change alters coastal marine environments around the world, the dynamics of local
43 recruitment, site fidelity and local adaptation are expected to change. This highlights the
44 importance of integrating knowledge about eco-evolutionary processes into marine resource
45 management, fisheries stock assessment, and restocking and stock enhancement activities.

46 **Keywords:** *ecological genomics, marine biogeography, teleost, recruitment dynamics,*
47 *fisheries management.*

48 **Introduction**

49 Understanding how evolutionary processes such as gene flow and local adaptation shape the
50 distribution of marine populations are enduring challenges in evolutionary biology and for
51 fisheries management (Bernatchez et al., 2017; Grummer et al., 2019). This is particularly
52 true for many marine species characterised by large population sizes, high fecundity and high
53 connectivity (Gagnaire et al., 2015), including those where recruitment dynamics is
54 influenced by sweepstakes reproductive success (Árnason et al., 2024). In these cases,
55 extensive gene flow could limit local adaptation, even across vast geographical ranges
56 (Lenormand, 2002; Nielsen, Hemmer-Hansen, Larsen, & Bekkevold, 2009). Despite these
57 challenges, growing evidence indicates that locally adapted genetic variation can be
58 maintained in species with high gene flow (Han et al., 2020).

59 Marine fishes provide ideal natural systems to examine the genomic basis of local adaptation
60 with gene flow, particularly in the context of fisheries resources. The management
61 consequences of local adaptation for widespread fisheries stocks are, however, relatively
62 unexplored (Andersson et al., 2024; Grummer et al., 2019). Highly dispersive marine species
63 often span extremely heterogeneous environments, yet conventional stock assessments may
64 fail to capture this complexity. Additionally, fisheries management and stock assessments are
65 often constrained by government jurisdictional boundaries that reflect political or
66 administrative divisions rather than biologically- or ecologically-relevant population structure
67 (Andersson et al., 2024). This potential disparity between management units and biological
68 populations can lead to localized overfishing, inadequate protection of important spawning or
69 nursery areas, or failure to account for source-sink dynamics occurring at larger spatial scales
70 (Berger et al., 2021).

71 Seascape genomics offers a powerful approach to disentangle demographic and ecological
72 evolutionary processes in these systems (Gagnaire et al., 2015). This interdisciplinary field
73 can clarify ecological and evolutionary factors shaping population structure and connectivity
74 of widespread fisheries species (Xuereb, d'Aloia, Andreollo, Bernatchez, & Fortin, 2021). By
75 examining how the environment influences genetic variation and gene flow, seascape
76 genomics can reveal subtle population structure that may be overlooked by previous genetic
77 methods or stock assessment approaches (Sandoval & Castillo, Robinson, Hart, Strain, &
78 Beheregaray, 2018). Atlantic herring (*Clupea harengus*) provide a clear example where,
79 despite limited evidence for population structure at neutral loci, seascape genomics revealed
80 fine-scale differentiation associated with ecological adaptation that helped to resolve
81 mismatches between biological populations and fisheries stock management (Han et al.,
82 2020). Similarly, temperature-associated adaptive variation was identified in the
83 commercially important yellow croaker (*Larimichthys crocea*) along the Chinese coast,
84 challenging the existing stock assessment and highlighting the need for more nuanced
85 management strategies (Chen et al., 2023).

86 Coastal marine species often exhibit limited population structure across continental-scale
87 distributions even if they encompass a wide range of environments. If patterns of dispersal
88 are locally restricted or temporally variable within these large populations, environmental
89 heterogeneity can, however, create opportunities for natural selection to drive local
90 adaptation. For these reasons, spatially and temporally heterogeneous coastal habitats, such as
91 the zonal coastal boundary of southern Australia, provide ideal opportunities to test for the
92 role of ecologically divergent natural selection in driving adaptation in marine species
93 (Ruzzante et al., 2006; Sandoval & Castillo & Beheregaray, 2020).

94 Here we examine the roles of regional and fine-scale connectivity linked to variable
95 recruitment dynamics in shaping population connectivity and adaptation in the Australasian
96 snapper (*Chrysophrys auratus*), a highly mobile coastal marine species. The focal region of
97 our study spans a range of heterogeneous embayment and open coastal environments along
98 more than 1500 km of the southern Australian coast (Ridgway & Condie, 2004). Snapper
99 plays a pivotal role in the commercial and recreational fisheries sector of southern Australia,
100 as well as holding considerable ecological and cultural value across its Indo-Pacific
101 distribution (Parsons et al., 2014). Recent declines in the productivity and status of some
102 snapper stocks have prompted significant shifts in management practices, including an
103 ongoing moratorium, now in its sixth year, on commercial and recreational fishing in much of
104 South Australia (Drew et al., 2022; Fowler et al., 2020). To inform and assess management of
105 the species it is critically important to understand how connectivity and environmental
106 variation shape snapper stocks subject to both fishing pressure and climate change. Several
107 genetic studies have revealed evidence for fine-scale population structure and provide a basis
108 for the hypothesis that selection, imposed by steep environmental gradients, could counteract
109 high gene flow and lead to local adaptation of snapper populations (Bernal-Ramírez, Adcock,
110 Hauser, Carvalho, & Smith, 2003; Bertram et al., 2023; Gardner, Chaplin, Potter, Fairclough,
111 & Jackson, 2017). Spawning aggregations and nursery areas across southern Australia
112 concentrate in three major embayments: northern Spencer Gulf, northern Gulf St. Vincent,
113 and Port Phillip Bay. Previous genetic analyses suggest these populations exhibit local
114 recruitment and site fidelity (Bertram et al., 2023), traits that favour the evolution of local
115 adaptation. This is supported by early molecular evidence for local adaptation in other
116 embayment populations, including Shark Bay, Western Australia (Johnson, Creagh, &
117 Moran, 1986), and New Zealand (Smith, 1979; Smith & Francis, 1983). Adaptive traits such
118 as larval survival, growth and metabolism have been strongly linked to temperature and

119 salinity in snapper (Fielder, Bardsley, Allan, & Pankhurst, 2005; McMahon, Parsons,
120 Donelson, Pether, & Munday, 2020; Wellenreuther, Le Luyer, Cook, Ritchie, & Bernatchez,
121 2019), and genomic variation is known to underpin these traits (Ashton, Hilario, Jaksons,
122 Ritchie, & Wellenreuther, 2019a; Ashton, Ritchie, & Wellenreuther, 2019b; Sandoval-
123 Castillo, Beheregaray, & Wellenreuther, 2022).

124 In this study, we predicted that genomic variation should not only reflect demographic factors
125 such as large population sizes and high gene flow, but also natural selection in response to
126 environmental heterogeneity. To test these predictions, we built on recent evaluations of
127 snapper stock structure and population size estimates (Bertram et al., 2022; Bertram et al.,
128 2023; Bertram et al., 2024) and examined how environmental parameters may influence
129 patterns of genomic variation and connectivity in snapper populations along the southern
130 Australian coast. Considering the social, economic, and commercial importance of southern
131 Australian snapper stocks, we also aimed to assess the findings in the context of informing
132 and refining regional fisheries management strategies which are fundamental for sustainable
133 harvesting and species persistence.

134

135 **Methods**

136 *Southern Australian coastal environment*

137 The focal region of this study, southern Australia, comprises a wide range of coastal and
138 near-shore marine habitats and includes three major embayments. Spencer Gulf and the
139 adjacent Gulf St. Vincent are large inverse estuaries in South Australia, and Port Phillip Bay
140 is a shallow, enclosed bay just south of Melbourne, Victoria (Figure 1). Broadly,
141 environmental conditions within these embayments differ greatly from open coastal habitats,
142 with cooler minimum sea surface temperature (SST), warmer maximum SST and reduced

143 current velocities. A west-to-east temperature gradient characterises the region, with the
144 warmest waters along the west coast and coolest in the eastern embayments. Regional
145 variation in climate, topography and oceanography create unique local environments within
146 each embayment. Climate in the east is wetter and cooler than the more arid South Australia.
147 As a result, Port Phillip Bay and Western Port Bay exhibit the coolest SST and receive
148 greater freshwater inputs from rivers, leading to the highest primary productivity in the region
149 and consistently low salinity. In South Australia, Spencer Gulf is characterised by extreme
150 salinity gradients, particularly in the north where values exceed 45 ppt in summer, combined
151 with large seasonal variation in SST (~12–24°C) (Nunes & Lennon, 1986). Gulf St. Vincent
152 experiences intermediate conditions that are less saline than Spencer Gulf but saltier than
153 coastal waters. Coastal current circulation outside the Gulfs is dominated by the Leeuwin
154 current system that transports tropical water poleward along the west coast of Australia and
155 eastward along the southern continental shelf, before joining the South Australian Current,
156 and eventually heading south along Tasmania’s west coast as the Zeehan Current
157 (Richardson, Middleton, Kyser, James, & Opdyke, 2019; Ridgway & Condie, 2004). The
158 strength and impact of this current system is highly variable and shapes seasonal patterns of
159 temperature, salinity and primary productivity across the region. Stronger currents during
160 winter transport warm, low-salinity water to the east, while reduced Leeuwin current flows
161 and coastal upwellings incorporate cooler, more saline and nutrient-rich water during summer
162 (Richardson et al., 2019; Ridgway & Condie, 2004; Shute et al., 2022).

163

164 *Sampling and genomic data collection*

165 Our sampling design builds on previous studies that clarified neutral population genetic
166 structure and stock boundaries of snapper along the western (Bertram et al., 2022) and the
167 southeastern (Bertram et al., 2023; Bertram et al., 2024) coasts of Australia. A total of 448

168 snapper (*Chrysophrys auratus*) collected from 23 individual sampling sites along the
169 southern Australian coast were used. These sites were grouped into ten regional locations
170 based on proximity and similarity of environmental profiles. To ensure adequate sample sizes
171 for allele-frequency estimation, individual sites containing fewer than six samples were
172 merged with the nearest neighbour. This resulted in 16 pooled sites representing the ten
173 regional locations (Figure 1; Figure S1; Table S1). All the research complies with applicable
174 laws on sampling from natural populations. We extracted DNA from fin clips using a salting-
175 out method (Sunnucks & Hales 1996) and used double-digest restriction site-associated DNA
176 (ddRAD) sequencing to generate genomic data. We generated ddRAD libraries of 96
177 multiplexed samples, following the protocol outlined in Peterson et al. (2012) with
178 modifications described in our previous work (Brauer, Hammer, & Beheregaray, 2016).
179 Paired-end 150bp sequencing was conducted over 15 lanes (this included samples not related
180 to this study) of an Illumina HiSeq 4000 at Novogene (Hong Kong). Raw sequence data
181 quality was assessed using FastQC (Andrews, 2010). Reads were demultiplexed with the
182 process_radtags module from STACKS 2.0 (Catchen, Hohenlohe, Bassham, Amores, &
183 Cresko, 2013) and trimmed to remove low-quality bases and adapters using
184 TRIMMOMATIC (Bolger, Lohse, & Usadel, 2014). Reads were then mapped to a
185 chromosome-level genome assembly (Catanach et al., 2019) using BOWTIE 2 (Langmead &
186 Salzberg, 2012), and SNPs were called using BCFTOOLS (Narasimhan et al., 2016). We
187 filtered the resulting SNP genotypes for quality, missing data per individual, and retained one
188 SNP per 500bp. We also applied further filtering to generate putatively neutral and candidate
189 adaptive datasets. Candidate adaptive loci identified by the genotype–environment
190 association analysis (described below) were separated, and the remaining putatively neutral
191 SNPs were filtered for departure from Hardy–Weinberg equilibrium (FDR of 0.05) using the

192 gl.filter.hwe function in the *dartR* package. Filtering parameters and the number of SNPs
193 retained following each step are described in Table S2.

194

195 *Genetic diversity, population structure and connectivity*

196 We estimated population genetic diversity parameters, including the number of alleles,
197 percentage of polymorphic loci, heterozygosity, and inbreeding coefficient, using the
198 *hierfstat* R package (Goudet, 2005). We examined patterns of neutral population structure
199 and admixture using using *hierfstat* to estimate pairwise F_{ST} and ADMIXTURE (Alexander,
200 Novembre, & Lange, 2009). An analysis of molecular variance (AMOVA) was performed to
201 further assess hierarchical population structure among major sampled regions, among sites
202 within regions, among individuals within sites, and within individuals. The *poppr.amova*
203 function in the *poppr* R package (Kamvar, Tabima, & Grünwald, 2014) was used for the
204 molecular variance estimates and significance values were estimated using the *ade4* *randtest*
205 function (Dray & Dufour, 2007) with 1000 permutations. We quantified gene flow among
206 sites within each of the eastern and western regional populations with the directional relative-
207 migration (N_m) approach of Sundqvist (2016), as implemented by the *divMigrate* function in
208 the *diveRsity* R package (Keenan, McGinnity, Cross, Crozier, & Prodöhl, 2013). This
209 approach first calculates pairwise differentiation (G_{ST}), then converts these values into unit-
210 less estimates of relative migrants per generation and assigns directionality by contrasting
211 each population's allele frequencies with those of a hypothetical mixed migrant pool.
212 Analyses were run separately for the neutral and putatively adaptive SNP datasets, and
213 statistical support was evaluated with 1,000 bootstrap resamples.

214

215

216

217 *Local Adaptation*

218 Environmental heterogeneity across the study area was summarised with 31 raster layers at 5-
219 arc-min resolution (~9 km at the equator) obtained from BioORACLE v2.2 (Assis et al.,
220 2018; Tyberghein et al., 2012), supplemented by bathymetry from BioORACLE v1.0 (Table
221 S3). Each layer represents long-term summary statistics (e.g. mean, minimum, maximum, or
222 range), calculated over a baseline period of 2000-2014 and derived either from the ECMWF
223 ORAP5.0 ocean reanalysis (physical variables), or the PISCES biogeochemical hind-cast
224 (nutrients and productivity). Two variables rely on different archives and time periods, pH
225 (1910-2007 observations, World Ocean Database) and calcite (2002-2009 MODIS-Aqua).
226 We performed preliminary analyses using the *vifcor* and *vifstep* functions in the *usdm* R
227 package (Naimi, Hamm, Groen, Skidmore, & Toxopeus, 2014) to remove correlated (Pearson
228 $r > 0.7$) and colinear ($VIF > 10$) variables. The retained environmental variables were finally
229 converted to z-scores to standardize across different measurement scales and ensure
230 comparable contributions to downstream analyses.

231

232 To detect a genetic signal of local adaptation, we performed genotype-environment
233 association analyses using redundancy analysis (RDA) implemented with the *rda* function in
234 the *vegan* R package (Oksanen et al., 2018). To examine relationships between SNP allele
235 frequencies and environmental data, we ran an initial partial RDA where we controlled for
236 spatial population structure using a matrix of allele frequency covariance (Ω) estimated with
237 Baypass (Gautier, 2015). Candidate adaptive loci significantly associated with environmental
238 variation were identified using the Mahalanobis distance statistical approach proposed by
239 Capblancq et al. (2018). A second RDA was then performed using the candidate adaptive loci
240 and individual SNP genotypes to visualise the distribution of genotype-environment
241 associations within populations.

242

243 Functional annotations of candidate loci were performed using SnpEff to assess genomic
244 position and predicted effects (Cingolani et al., 2012). We then extracted open reading frames
245 (ORFs) from 600 bp flanking sequences around each SNP and searched these against the
246 SwissProt teleost database using DIAMOND BLASTx (Buchfink, Xie, & Huson, 2015),
247 applying an e-value threshold of 1×10^{-10} and retaining up to five hits per query. Top-
248 scoring matches were queried through the UniProt REST API to retrieve Gene Ontology
249 (GO) terms and KEGG pathways, retaining only those with experimental or curated evidence
250 codes (EXP, IDA, IPI, IMP, IGI, IEP, TAS, IC) or high-confidence computational inference
251 (ISS, IEA). To identify enriched functional categories, we performed over-representation
252 analysis using the *clusterprofiler* R package (Yu, Wang, Han, & He, 2012). Annotated
253 candidate loci were compared against all annotated loci using hypergeometric tests, with false
254 discovery rate controlled by the Benjamini–Hochberg procedure ($FDR < 0.1$). Categories
255 containing fewer than five or more than 500 genes were excluded to avoid unstable or overly
256 general terms. The genomic distribution of candidate loci and enriched functional categories
257 was visualized using the *ggmanh* R package (Lee, 2022).

258

259 **Results**

260 *Sampling and genomic data collection*

261 The Illumina HiSeq 4000 sequencing generated 5.05 billion raw sequences, and 3.36 billion
262 (mean per sample = 7.51M, min = 0.87M, max = 21.38M) were retained. After filtering, we
263 retained 448 individuals genotyped at 14,699 single nucleotide polymorphisms (SNPs).
264 Filtering for Hardy–Weinberg equilibrium (HWE), retained 14,206 SNPs, and exclusion of
265 candidate adaptive loci resulted in a putatively neutral dataset of 13,453 SNPs.

266

267 *Genetic diversity, population structure and connectivity*

268 Estimates of genetic diversity were based on the 13,453 neutral loci, with observed
269 heterozygosity (H_o) ranging from 0.124 in southern Gulf St. Vincent to 0.138 in southern
270 Spencer Gulf, while expected heterozygosity (H_e) varied from 0.128 to 0.133. Inbreeding
271 coefficient (F_{is}) values were generally low, ranging from -0.036 in southern Spencer Gulf to
272 0.036 in northern Gulf St. Vincent, indicating minimal inbreeding within populations.
273 Percentages of polymorphic loci ranged from 79.1% in Kingston–South East to 87.0% in Port
274 Phillip Bay (Table 1).

275

276 ADMIXTURE results indicated that the optimal number of genetic clusters was $K = 2$
277 (Figure 2), corresponding to an eastern and a western cluster with the genetic break situated
278 between southern Gulf St. Vincent and Kingston–South East. Despite this main division, a
279 few migrants (three from the eastern and two from the western cluster) are evident in the
280 ADMIXTURE plot. Pairwise F_{ST} values based on the 13,453 neutral loci were generally low
281 among sites within regions but higher between eastern and western regions (mean = 0.009,
282 range = 0–0.018). Based on the candidate adaptive loci, F_{ST} estimates were elevated, both
283 among sites within each region, and among sites across regions (mean = 0.013, range = 0–
284 0.25; Figure S2). The AMOVA analyses supported the ADMIXTURE results, attributing a
285 significant proportion of genetic variation to differences between regions (2.1%,
286 $\Phi_{CT} = 0.021$, $P < 0.01$), while variation among sites within regions was minimal (0.1%,
287 $\Phi_{SC} = 0.001$, $P < 0.001$). The AMOVA based on candidate adaptive loci assigned a higher
288 proportion of variation between sites within regions (0.4%, $\Phi_{SC} = 0.004$, $P < 0.001$), and
289 among regions (5.9%, $\Phi_{CT} = 0.059$, $P < 0.01$), indicating stronger differentiation at loci
290 potentially under selection (Table 2).

291

292 The relative migration (N_m) estimates based on neutral loci for both the eastern and western
293 regions showed high connectivity among sites (eastern mean $N_m = 0.79$, western mean $N_m =$
294 0.77 ; Figure 3a,c, Table S4-5), with particularly high connectivity evident between the
295 southern areas of the two gulfs, Spencer Gulf and Gulf St. Vincent (mean $N_m = 0.97$), and
296 between Port Phillip Bay and all other eastern sites (mean $N_m = 0.89$). Using the candidate
297 loci, migration estimates for eastern region were lower (mean $N_m = 0.72$; Figure 3b, Table
298 S6). For the western region, relative migration estimates again suggested the southern areas
299 of both gulfs are highly connected (mean $N_m = 0.98$; Figure 3d, Table S7) but indicated
300 much lower connectivity for all pairwise estimates involving West Coast (mean $N_m = 0.53$)
301 and, particularly northern Spencer Gulf (mean $N_m = 0.46$).

302

303 Filtering correlated (Pearson $r > 0.7$) and colinear ($VIF > 10$) variables resulted in a final set
304 of five environmental variables describing environmental heterogeneity across the study
305 region (minimum sea surface temperature, mean sea surface salinity, minimum sea surface
306 net primary productivity, pH, and calcite concentration; Figure S3, Table S8). These variables
307 capture critical factors known to influence snapper physiology and life history, including
308 thermal tolerance during winter months, osmoregulatory stress in hypersaline embayments,
309 food availability during low-productivity periods, and carbonate chemistry affecting
310 calcification processes. The partial RDA identified 855 candidate adaptive loci (5.8% of the
311 total SNPs) significantly associated with the environment (Figure 4a). The individual-level
312 RDA based on those 855 candidate loci (Figure 4b) revealed divergent genotype-environment
313 associations between regions, with western populations showing strong associations with
314 salinity and calcite (reflecting the hypersaline Spencer Gulf environment), while eastern
315 populations were more strongly associated with temperature minima and productivity
316 gradients.

317

318 Functional annotation of these loci revealed a substantial proportion located in genic regions,
319 with many predicted to have moderate to high impacts on gene function (Figure 5a-b). We
320 recovered 1,188 BLAST hits covering 730 unique loci (85.4% of candidates). These matches
321 corresponded to 562 unique proteins with 5,115 GO terms (2,640 biological process, 1,263
322 cellular component, and 1,212 molecular function terms) and 488 KEGG pathway
323 annotations (Table S9). Based on these annotations, over-representation analysis identified
324 two enriched biological process GO terms. Post-anal tail morphogenesis (GO:0036342; $p =$
325 0.0022, FDR = 0.062), was represented by three genes involved in FGF-BMP-Wnt signaling
326 (*fgfr1a*, LG10; *tll1*, LG15; *bcl9l*, LG18), a conserved growth-factor network that regulates
327 skeletal and muscle growth and mediates plastic responses to environmental cues such as
328 temperature and salinity (Johnston, 2006). Hemopoiesis (GO:0030097) was also significantly
329 enriched ($p = 0.0043$, FDR = 0.062), with three genes affecting red blood cell production or
330 membrane composition (*rnf145*, LG16; *zfpml*, LG4; *smarcal1*, LG8). No significant
331 enrichment was detected for molecular function or cellular component categories, or for
332 KEGG pathways. The functionally annotated candidate loci were distributed across the
333 genome, with enriched genes found on multiple linkage groups (Figure 5c).

334

335 **Discussion**

336 Understanding how environmental heterogeneity shapes population structure and
337 connectivity remains a central challenge in ecology and evolution, particularly for marine
338 species with highly variable recruitment dynamics and dispersal potential. The relationship
339 between gene flow and local adaptation is especially complex in marine environments where
340 oceanographic connectivity can span vast distances while environmental gradients create
341 strong selective pressures. Along the southern Australian coast, our findings reveal that broad

342 environmental gradients influence patterns of dispersal and local adaptation in snapper,
343 despite substantial gene flow among populations. We identified two distinct regional
344 populations with minimal genetic differentiation at neutral loci among sites within each
345 region, consistent with high demographic connectivity. However, genotype-environment
346 association analyses identified 855 candidate adaptive loci linked to five key environmental
347 variables that shape snapper ecology. Minimum temperature that affects spawning, growth
348 and survival, salinity extremes challenging osmoregulation, primary productivity determining
349 prey availability, and carbonate chemistry (pH and calcite) potentially influencing sensory
350 systems and otolith formation (Cook, Herbert, & Jerrett, 2021; Fielder et al., 2005). A
351 substantial proportion of these putatively adaptive SNPs are located in genic regions with
352 potential moderate to high impacts on gene function. Functional over-representation analyses
353 identified six candidate SNPs that mapped to two enriched GO biological process terms
354 (post-anal tail morphogenesis; hemopoiesis), suggesting hydrodynamic niche and salinity
355 tolerance as potential targets of selection. While relative migration estimates using neutral
356 loci suggested high connectivity among sites within each population, using candidate loci
357 revealed lower connectivity, particularly for West Coast and northern Spencer Gulf
358 populations. This decoupling of demographic and adaptive connectivity highlights that
359 substantial gene flow does not preclude local adaptation, with specific environmental
360 stressors contributing to adaptive genetic divergence of local snapper populations.

361

362 *Strong environmental gradients influence connectivity and local adaptation*

363 Current knowledge of snapper population dynamics across southern Australia, based on size
364 structure, mark-recapture, and otolith studies, indicates that spatial and temporal patterns in
365 regional connectivity mainly reflect interannual variation in recruitment to three main nursery
366 areas: Port Phillip Bay in the east, and northern Spencer Gulf and northern Gulf St. Vincent

367 in the west (Drew et al., 2022; Fowler et al., 2020; Hamer, Jenkins, & Gillanders, 2003). We
368 detected evidence for occasional long-distance migration from the eastern stock into southern
369 Gulf St. Vincent, along with more subtle, fine-scale differentiation between sub-populations
370 within, and adjacent to the South Australian gulfs. The major genetic break we observed
371 between the eastern and western populations aligns with the boundary between the well-
372 described Flindersian and Maugean marine biogeographic provinces, suggesting that broad-
373 scale climatic and oceanographic processes are important drivers of stock structure across the
374 region (Teske, Sandoval-Castillo, Waters, & Beheregaray, 2017; Waters et al., 2010). These
375 findings are consistent with previous genetic studies (Bertram et al., 2023), and with the view
376 that strong recruitment events drive dispersal of juvenile snapper among populations at both
377 local and regional spatial scales (Fowler, 2016).

378

379 Although the coast of southern Australia is relatively well-studied from a biogeographic
380 perspective (Teske et al., 2017; Waters, 2008), less is known about how its heterogeneous
381 seascape and strong environmental gradients influence connectivity and local adaptation,
382 particularly for teleosts. Seasonal variation in the intensity and direction of prevailing
383 oceanographic currents can shape local environmental conditions including temperature,
384 salinity and dissolved oxygen (Richardson et al., 2019). Such variation is also known to
385 mediate evolutionary processes such as dispersal, recruitment, and local adaptation for a
386 range of coastal marine species including macroalgae, sea urchins, silversides, sardines, and
387 cetaceans (Banks et al., 2007; Barceló et al., 2022; Beheregaray & Sunnucks 2001; Coleman
388 et al., 2011; Teske, Sandoval-Castillo, Van Sebille, Waters, & Beheregaray, 2016; Teske et
389 al., 2021; Ward et al., 2006). For snapper, strong recruitment events can enhance
390 demographic connectivity at both regional and local scales, from long-distance westward
391 dispersal originating from the eastern nursery area (Fowler, 2016; Hamer & Jenkins, 2004) to

392 more localised exchange among subpopulations within regions. However, environmental
393 heterogeneity among nursery areas, and between embayments and adjacent coastal waters,
394 likely imposes strong selection on migrants that may favour survival of local recruits (Rankin
395 & Sponaugle, 2011). These patterns are consistent with a migration-selection balance, where
396 exceptional recruitment pulses are offset by selection against maladapted immigrants,
397 maintaining local adaptation across environmental gradients (Yeaman & Whitlock, 2011).
398
399 Supporting this hypothesis, steep environmental gradients between upper gulf and open
400 coastal environments corresponded with allele frequency differences at candidate adaptive
401 loci. This included strong associations with temperature and salinity that are known to impact
402 growth and survival of snapper (Fielder et al., 2005; McMahon et al., 2020). Previous
403 snapper studies have demonstrated that adaptive traits such as growth and survival are
404 strongly linked to these environmental variables (Wellenreuther et al., 2019), and have
405 identified genomic variation underpinning these traits (Ashton et al., 2019a; Ashton et al.,
406 2019b; Sandoval-Castillo et al., 2022). We found post-anal tail morphogenesis genes (*fgfr1a*,
407 *ill1*, *bcl9l*) and haemopoietic regulators (*zfpml*, *rnfl45*, *smarcal1*) were significantly over-
408 represented in our candidate adaptive loci. These functional modules align with growth-factor
409 and oxygen-transport pathways that Wellenreuther et al. (2019) showed were sensitive to
410 temperature in snapper, and have also been implicated in osmotic-stress responses in other
411 fishes (Fiol & Kültz, 2007). This suggests that the same canonical signalling networks could
412 mediate adaptation to the joint temperature-salinity gradients that span the embayment-
413 coastal habitats. More broadly, the distribution of candidate adaptive loci across many
414 regions of the genome is consistent with a signal of polygenic adaptation (Stephan, 2016).
415 While similar polygenic signals of selection have been observed in many marine species
416 (Bernatchez, 2016), it is important to note that alternative genomic architectures have also

417 been observed in high gene flow marine fishes. In Atlantic silversides, strong divergent
418 selection was shown to concentrate adaptive alleles into large genomic haploblocks (or
419 “supergenes”) maintained by suppressed recombination, despite an overall low level of
420 genome-wide differentiation (Wilder, Palumbi, Conover, & Therkildsen, 2020). Structural
421 variants can contribute to complex trait variation and can also create localised peaks of
422 differentiation. Recent whole-genome analyses in snapper revealed substantial structural
423 variation (Blommaert, Sandoval-Castillo, Beheregaray, & Wellenreuther, 2024) suggesting
424 that future genomic studies may yield additional adaptive features beyond those our ddRAD
425 approach could detect.

426

427 The role of natural selection in structuring marine populations across environmental gradients
428 is gaining appreciation and this understanding is crucial for predicting climate-driven changes
429 in fisheries sustainability (Teske et al., 2021). Local adaptation to temperature has been
430 documented in economically important species such as the large yellow croaker
431 (*Larimichthys crocea*), where genetic differentiation tracks minimum temperatures despite
432 high connectivity, resulting in climate-induced shifts in stock boundaries (Chen et al., 2023).
433 Similarly, salinity gradients were found to drive adaptive divergence in osmoregulation and
434 reproductive traits among sand goby (*Pomatoschistus minutus*) populations across the North
435 and Baltic Seas (Leder et al., 2021). Within Spencer Gulf and Gulf St. Vincent, where both
436 temperature and salinity increase to the north, these environmental gradients influence
437 multiple species. King George whiting (*Sillaginodes punctatus*), maintain genetic
438 homogeneity across the South Australian region, yet comprise ecologically independent
439 populations supplied by separate spawning grounds (Rogers, Fowler, Steer, & Gillanders,
440 2019). Despite their much longer pelagic larval phase (~100 days, compared with 3-4 weeks
441 in snapper), whiting exhibit the same fine-scale genetic structure linked to environmental

442 variation at spawning grounds and to distinct larval dispersal routes. The recurrence of this
443 pattern in taxa with contrasting life-history traits suggests a general pattern of environmental
444 heterogeneity shaping fine-scale connectivity and population structure in marine fish
445 populations across the region.

446

447 *Contingent migration can reinforce local adaptation with gene flow*

448 Local adaptation across environmental gradients despite high gene flow has been documented
449 in numerous marine species, including Atlantic cod (*Gadus morhua*), Pacific herring (*Clupea*
450 *harengus*), and sardines (*Sardinops sagax*) (Bradbury et al., 2010; Limborg et al., 2012;
451 Teske et al., 2021). These studies suggest that the balance between selection and gene flow in
452 marine environments may be more nuanced than often assumed. Contingent migration, where
453 populations contain both migratory and resident individuals, provides a potential general
454 mechanism to explain the observed balance between connectivity and local adaptation
455 (Hansson & Åkesson, 2014; Secor, 1999). Contingent migration strategies have been
456 described for several marine fishes, including southern flounder (*Paralichthys lethostigma*) in
457 the Gulf of Mexico (Steffen et al., 2023), and mulloway (*Argyrosomus japonicus*) and
458 southern garfish (*Hyporhamphus melanochir*) in Australia (Hughes, Meadows, Stewart,
459 Booth, & Fowler, 2022; Steer & Fowler, 2015). For snapper, tagging studies of the same
460 population have revealed complex movement patterns with some individuals showing high
461 site fidelity while others undertake extensive migrations (Fowler, 2016; Parsons et al., 2003;
462 Stewart, Pidd, Fowler, & Sumpton, 2019). This is supported by evidence for three distinct
463 behavioural groups in Port Phillip Bay snapper: fish that depart immediately after spawning,
464 summer residents that remain until autumn, and individuals that overwinter in the bay (Hamer
465 & Mills, 2017). Such behavioural variation is consistent with contingent migration where

466 migratory individuals facilitate gene flow, while resident contingents maintain locally
467 adapted genetic variation.

468

469 This balance between gene flow and selection is however unlikely to be temporally stable.

470 The relative proportions of resident versus migratory contingents can shift in response to

471 environmental variability and recruitment pulses, as documented in white perch (*Morone*

472 *americana*) and striped bass (*M. saxatilis*) where contingent behaviour varied with river flow,

473 productivity, and climatic conditions (Gahagan, Fox, & Secor, 2015; Gallagher, Piccoli, &

474 Secor, 2018). For snapper, strong year classes might disperse more widely to reduce intra-

475 specific competition, while weaker year classes may reinforce local residency (Drew et al.,

476 2022). Previous genomic studies have detected temporal shifts in genetic composition,

477 reflecting these variable recruitment and connectivity dynamics (Bertram et al., 2023).

478 Complex patterns of snapper stock structure observed in Shark Bay, Western Australia,

479 further illustrate this temporal variation. Early tagging, otolith, and allozyme studies detected

480 strong isolation between gulf and coastal populations (Jackson & Moran, 2012; Johnson et

481 al., 1986), however subsequent microsatellite work revealed more subtle differentiation

482 linked to salinity and timing of spawning (Gardner et al., 2017). These dynamics may also

483 change under climate warming, potentially altering the timing and intensity of episodic

484 recruitment or shifting the strength of environmental selection across heterogeneous

485 spawning habitats. This highlights the importance of integrating knowledge about ecological

486 and evolutionary processes into fisheries stock assessment and management, and into

487 restocking and stock enhancement programs aimed at improving the abundance and

488 resilience of wild stocks (e.g. Harrison et al., 2025).

489

490 Our study provides novel insights into the complex relationship between gene flow and local
491 adaptation in snapper populations along southern Australia's environmentally heterogeneous
492 coastline. The identification of genomic regions potentially under selection reveals restricted
493 gene flow at candidate adaptive loci, highlighting that environmental factors shape population
494 structure even in highly connected species like snapper. The potential role of contingent
495 migration, where migratory and resident individuals coexist within populations, appears to be
496 a key mechanism facilitating the balance between connectivity and local adaptation. This has
497 direct implications for fisheries management in the region, particularly given the recent
498 decline of some stocks. While current management necessarily operates within practical
499 jurisdictional frameworks, our findings reveal biologically and ecologically relevant
500 population structure that could enhance existing frameworks by incorporating both the
501 demographic connectivity between regions and the adaptive distinctiveness of local
502 populations. For instance, the Spencer Gulf and West Coast populations, currently managed
503 as a single stock, show distinct adaptive signatures that may warrant consideration in refining
504 future management strategies. Preserving these locally adapted populations that contribute to
505 regional recruitment may prove crucial for persistence of snapper and other coastal species
506 under climate change. While temporal variability in recruitment is already understood,
507 longitudinal genomic monitoring could enhance assessment frameworks by revealing how
508 this variability affects connectivity patterns and adaptive potential. Furthermore,
509 strengthening collaboration between management jurisdictions would help ensure that
510 population dynamics operating at large spatial scales are captured within local management
511 strategies. As fisheries continue to experience unprecedented environmental changes,
512 management strategies that integrate knowledge of both neutral and adaptive genetic
513 variation, as well as of adaptive capacity, will be essential for ensuring long-term
514 sustainability. Our study shows that demographic and adaptive connectivity can be decoupled

515 in highly connected marine species and highlights that cryptic adaptive variation may prove

516 crucial for population persistence in changing oceans.

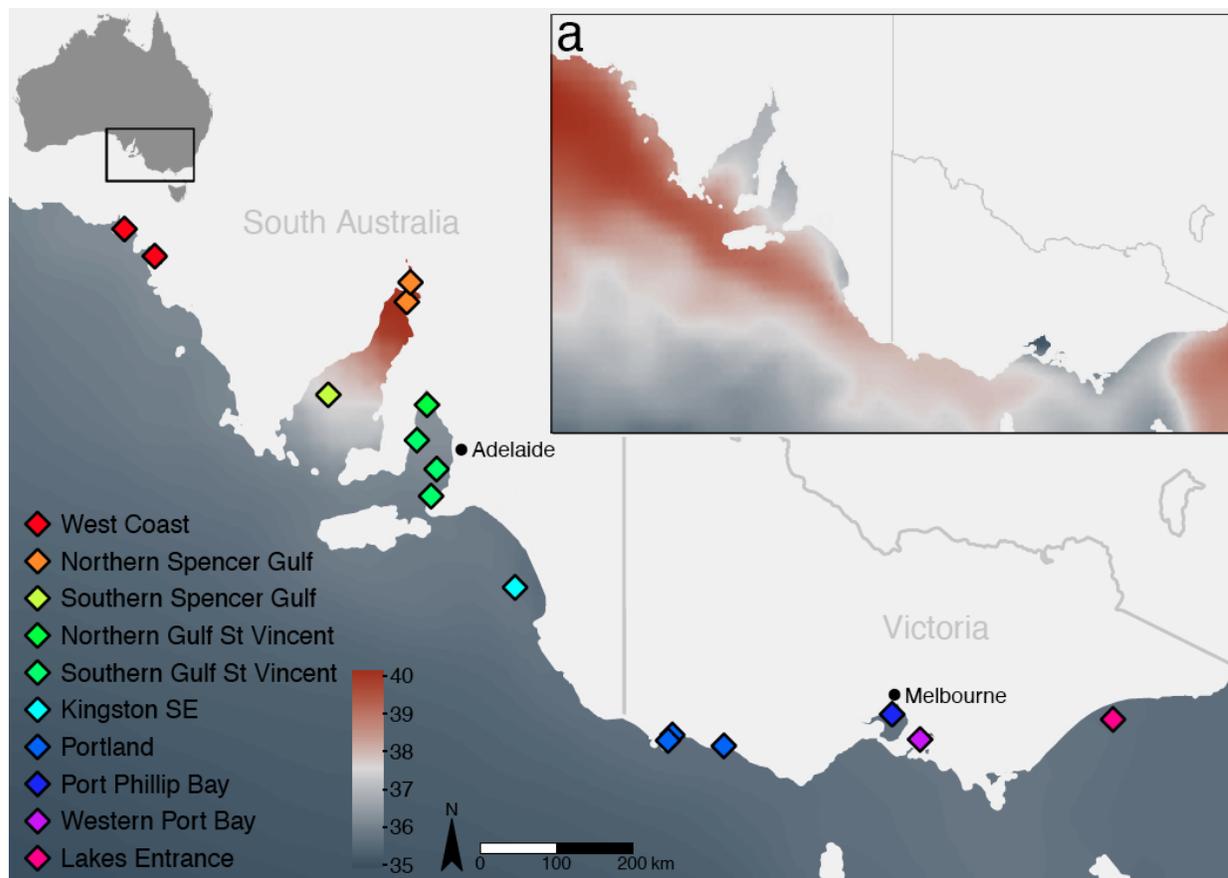
517

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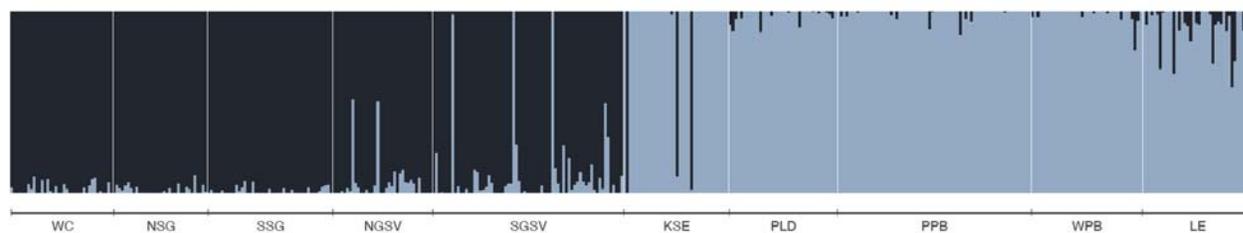


528

529 Figure 1. Sampling sites across southern Australia and spatial heterogeneity in mean sea
530 surface salinity (main panel), and (a) minimum sea surface temperature.

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532



533

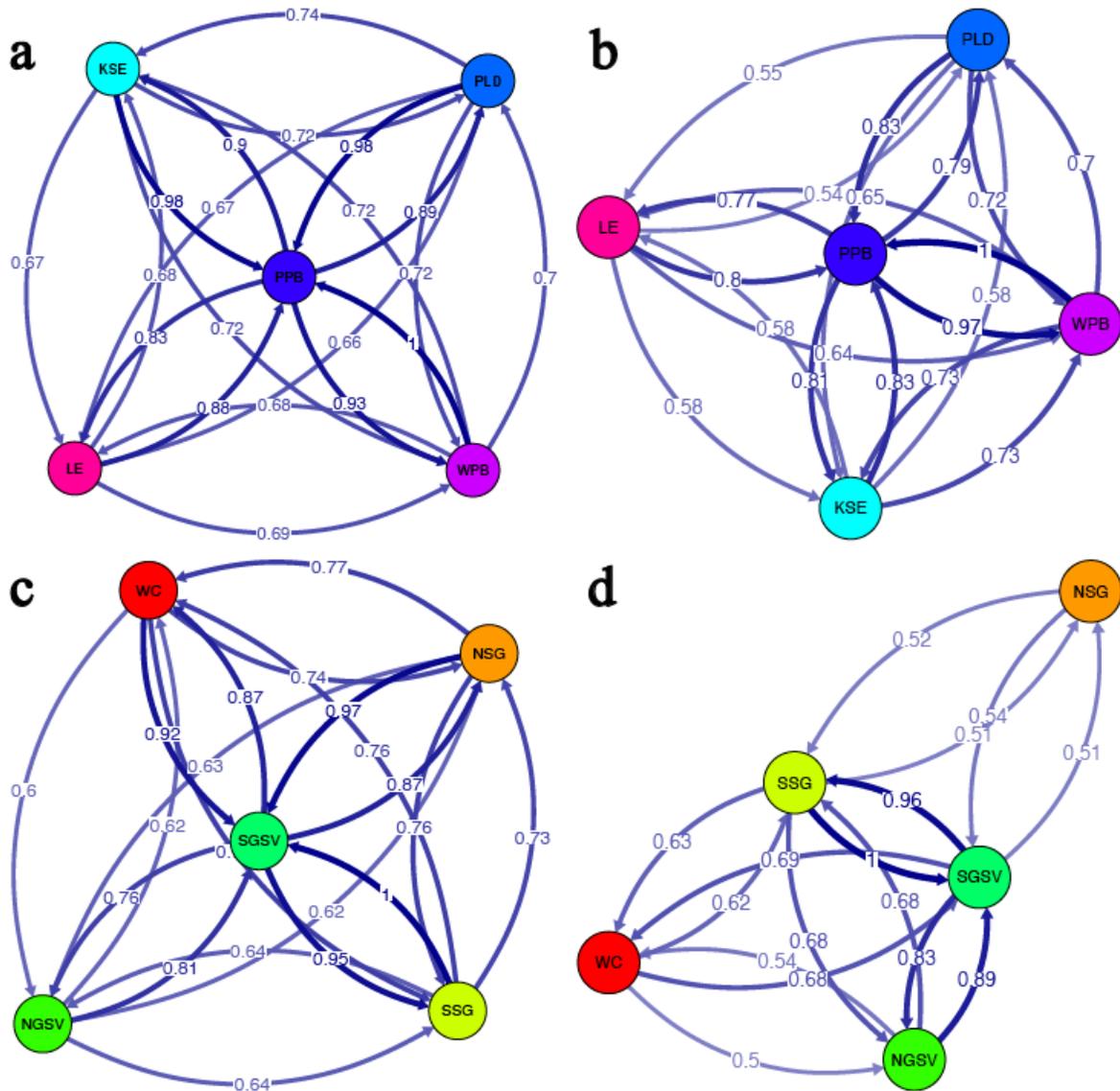
534 Figure 2. ADMIXTURE results for K=2, indicating distinct western (dark) and eastern

535 (light) genetic clusters. Site codes refer to the locations listed in Table 1.

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540 Figure 3. Relative migration networks for the eastern stock estimated using (a) 13,453 neutral

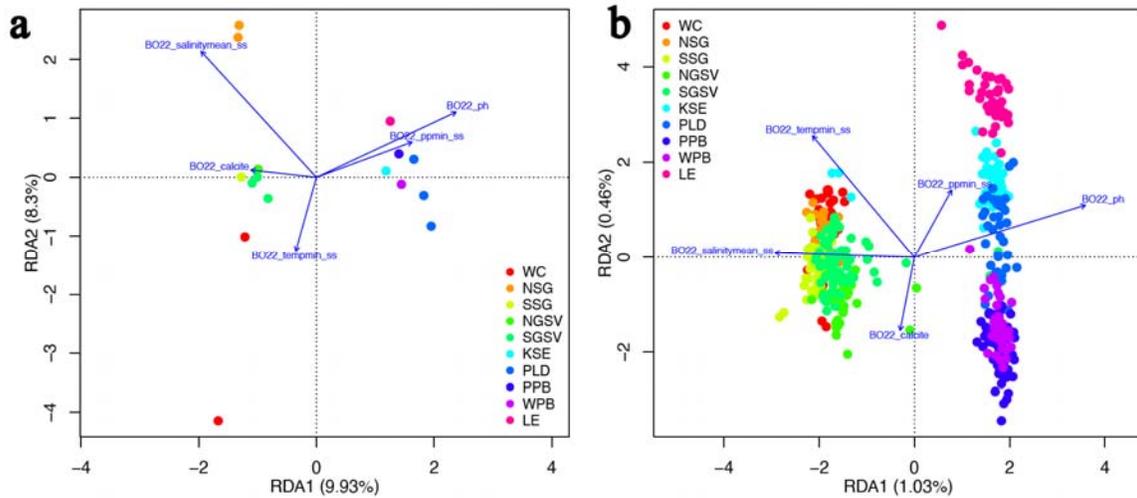
541 and (b) 855 candidate adaptive SNPs, and for the western stock using (c) 13,453 neutral and

542 (d) 855 candidate SNPs, following the method of Sundqvist (2016). Darker arrows signify

543 higher relative migration. Estimates <0.5 are not shown.

544

545



546

547

548 Figure 4. Redundancy analysis (RDA) plots summarising genotype–environment associations

549 in Australasian snapper: (a) RDA biplot showing ordination of sampling locations based on

550 14,699 genome-wide SNPs after controlling for spatial structure using a population allele

551 frequency covariance matrix. Arrows indicate the direction and strength of environmental

552 gradients, including minimum sea surface temperature, mean sea surface salinity, minimum

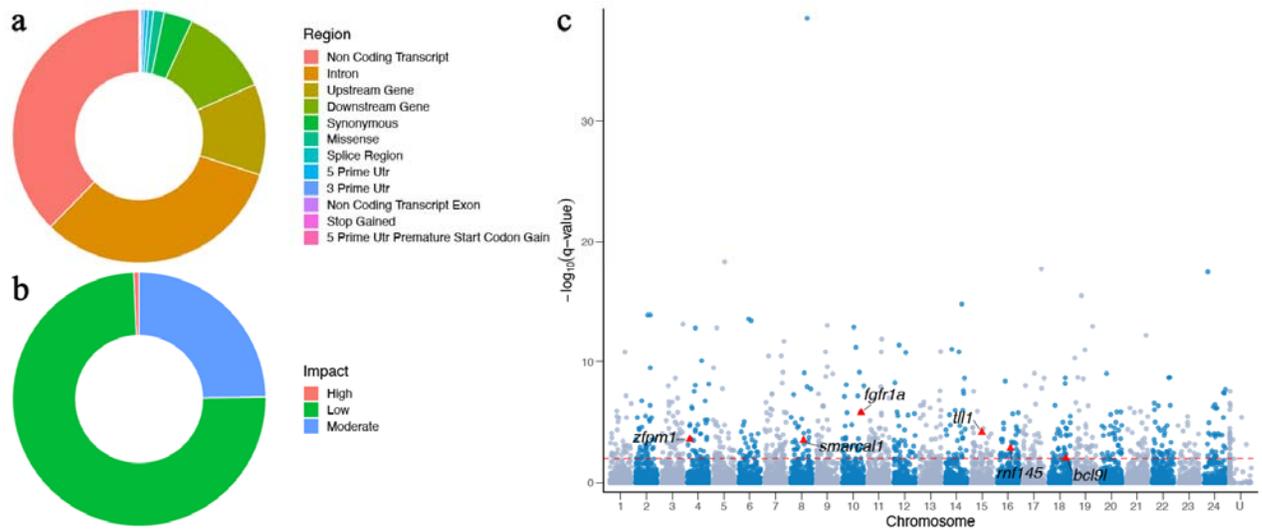
553 sea surface net primary productivity, pH, and calcite concentration; (b) Individual-level RDA

554 biplot based on 855 putative adaptive candidate loci, illustrating the distribution of individual

555 genotypes across sites in relation to environmental variables.

556

557



558

559 Figure 5. (a) Proportion of 855 candidate adaptive SNPs by genomic region; (b) Proportion of
560 candidate SNPs in genic regions by impact on gene function; (c) Manhattan plot indicating
561 the RDA q-values ($-\log_{10}[q]$) and distribution of 855 candidate adaptive SNPs associated with
562 environmental variation (FDR 0.01; above dashed line). Red triangles highlight loci linked to
563 six genes representing two functional gene ontology terms enriched in the candidate adaptive
564 SNPs (FDR < 0.1), GO:0036342, post-anal tail morphogenesis (*fgfr1a*, *tll1*, *bcl9l*), and
565 GO:0030097, hemopoiesis (*rnf145*, *zfp1*, *smarca1*).

566

567 Table 1. Genetic diversity indices based on 13,453 neutral loci. Number of individuals, N;
 568 number of alleles, N_A ; percentage polymorphic loci, %PL; observed heterozygosity, H_O ;
 569 expected heterozygosity, H_E ; and inbreeding coefficient, F_{IS} (including 95% confidence
 570 intervals, LCI and UCI).

Site	N	N_A	%poly	H_O	H_E	F_{IS}	LCI	UCI
West Coast (WC)	38	1.610	83.5	0.125	0.129	0.033	0.028	0.036
Northern Spencer Gulf (NSG)	40	1.607	83.3	0.125	0.128	0.019	0.016	0.023
Southern Spencer Gulf (SSG)	36	1.625	84.2	0.138	0.133	-0.036	-0.040	-0.032
Northern Gulf St Vincent (NGSV)	34	1.626	83.4	0.127	0.131	0.036	0.032	0.041
Southern Gulf St Vincent (SGSV)	39	1.597	80.7	0.124	0.128	0.030	0.027	0.034
Kingston SE (KSE)	70	1.602	79.1	0.128	0.130	0.013	0.009	0.016
Portland (PLD)	69	1.633	85.6	0.129	0.132	0.022	0.019	0.026
Port Phillip Bay (PPB)	45	1.623	87.0	0.129	0.131	0.013	0.008	0.017
Western Port Bay (WPB)	37	1.628	85.1	0.131	0.132	0.010	0.006	0.014
Lakes Entrance (LE)	40	1.601	81.8	0.129	0.129	0.000	-0.004	0.005

571

572

573 Table 2. Hierarchical analysis of molecular variance (AMOVA) based on 13,453 neutral and
 574 855 candidate loci, illustrating the distribution of genetic variation among regions (eastern,
 575 western), sites within each region, and individuals within sites, and within individuals.

SNPs	Source of variation	df	Sum of squares	Sigma (σ)	% variance	Phi (Φ)	P-value
13,453	Among regions	1	10590.788	21.112	2.1	0.021	0.009
	Among sites within regions	8	8988.927	1.184	0.1	0.001	0.001
	Among individuals within sites	438	446601.017	13.373	1.3	0.013	0.035
	Within individuals	448	444815.000	992.891	96.5	0.035	0.001
855	Among regions	1	4689.312	9.958	5.9	0.059	0.008
	Among sites within regions	8	1785.722	0.630	0.4	0.004	0.001
	Among individuals within sites	438	73553.296	9.993	5.9	0.063	0.001
	Within individuals	448	66279.000	147.944	87.8	0.122	0.001

576

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869 **Declaration of Interest Statement**

870 The authors have nothing to declare.

871

872 **Data Accessibility and Benefit-Sharing**

873 The study datasets are available in *Figshare*:

874 <https://figshare.com/s/034cb8ca31701cf9b687>

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877 Benefits from this research accrue from the sharing of our data and results on public

878 databases as described above.

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